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## Characterization of Controlled Release Ofloxacin Suspensions by Fourier Transform Infrared Spectroscopy

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### ABSTRACT

Ofloxacin is having low solubility in aqueous solution and high rate of absorption from the stomach. It is precipitated at alkaline pH, leading to erratic absorption of the drug from small intestine. Moreover, its biological half-life is from 5 to 6 h. To overcome these difficulties, controlled release mucoadhesive suspensions have been designed so that safe and effective blood level of Ofloxacin can be maintained for a prolonged period. The chemical interaction between Ofloxacin and different polymers (Carbopol934, Carbopol940 and Hydroxypropyl methyl cellulose) in suspensions has been studied to know their compatibility by Fourier Transform Infrared Spectroscopy. From the spectral interpretation, it was found that in formulations, the carboxylic groups of Ofloxacin and hydroxyl groups of respective polymers encountered chemical interaction, leading to esterification and hydrogen bonding (both intermolecular and polymeric). It may be concluded that Ofloxacin is compatible with three polymers used in the present study. Formation of micellies due to esterification and intermolecular hydrogen bonding causes more drug entrapment. In addition, stable suspensions are formed without hampering the C-F bond of the quinolone nucleus, which is responsible for the antibacterial activity of the drug. As a result, these polymers may be considered as effective carriers for Ofloxacin.

**Keywords:** Ofloxacin, C934, C940, HPMC, FTIR, Mucoadhesive Suspensions

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## INTRODUCTION

Ofloxacin (Oflox), 9-fluoro-2, 3-dihydro-3-methyl-10-(4-methyl-1-piperiziny)-7-oxo-7H-pyrido [1,2,3-de]-1,4-benzoxaine-6-carboxylic acid, is a fluoroquinolone antibacterial agent (Figure 1). Normal dosage regimen varies from 200 to 600 mg administered twice or thrice a day, depending on severity of infection. In severe cases, long-term therapy may also be required. Biological half-life of the drug is from 5 to 6 h. As frequent dosing is required to maintain the therapeutic plasma concentration, it was chosen as a model drug for the controlled release study [1]. Taking into consideration of above factors, polymeric suspensions of Ofloxacin were prepared by using two grades of mucoadhesive biodegradable carbopol polymers i.e., Carbopol934 (C934) and Carbopol940 (C940); and Hydroxypropyl methylcellulose (HPMC). This was done to protect the drug from the physiological environment, leading to improvement in its stability *in vivo*.

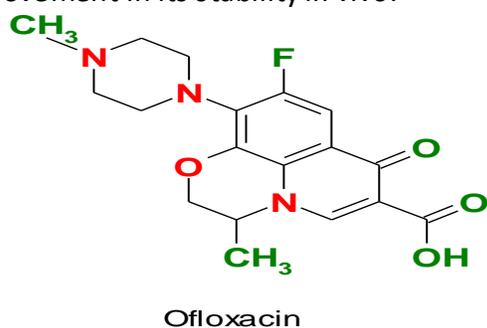


Figure 1: Chemical structure of Ofloxacin

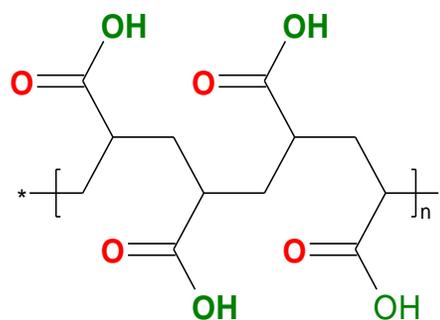


Figure 2: Chemical Structure of Carbopol Polymer

Both C934 and C940 consist of chains of polyacrylic acid (Figure 2) and they differ by the cross linking agents like allyl ethers of sucrose in C934 and allyl ethers of pentaerythritol in C940 [2,3]. Carbopol polymers are pH sensitive [4, 5] environmentally responsive polymer or considered as smart gels [6]. They have recently attracted considerable interest in the field of drug delivery as a means of providing an on-off release by shrinking and swelling in response to the change in pH [7-10].

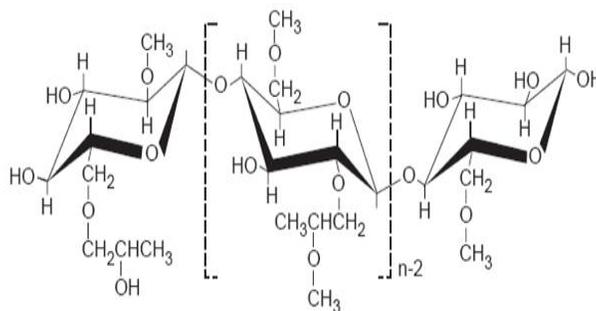


Figure 3: Chemical structure of Hydroxypropyl methylcellulose

Hydroxypropyl methylcellulose (HPMC) is propylene glycol ether of methyl-cellulose. Its chemical structure has been illustrated in Figure 3 [11]. It is one of the most commonly used hydrophilic biodegradable polymers for developing controlled release formulations, because it works as a pH-independent gelling agent. Swelling as well as erosion of it occurs simultaneously

inducing a pseudofed state, thereby reducing peristaltic contraction, which contributes to overall drug release. It is a widely accepted pharmaceutical excipient because HPMC is available in a wide range of molecular weights and the effective control of gel viscosity is easily possible [12-16]. It has many pharmaceutical uses, such as a drug carrier, a coating agent, a tableting agent, etc [11]. It is the most important hydrophilic carrier material used for the preparation of oral controlled drug delivery systems. One of its most important characteristics is the high swellability, which has a significant effect on the release kinetics of an incorporated drug. Upon contact with water or biological fluid, the latter diffuses into the device, resulting in polymer chain relaxation with volume expansion. Subsequently, the incorporated drug diffuses out of the system. Moreover, the physicochemical properties of HPMC are strongly affected by: (i) the methoxy group content; (ii) the hydroxypropoxy group content; and (iii) the molecular weight [12]. It may form a complex with the low solubility drug like Ofloxacin.

Since the information that can be provided by FTIR is identification of unknown materials along with quality, purity and consistency of the sample along with chemical interaction with other compounds [17, 18], to know the different functional groups and highly polar bonds of pure Ofloxacin and different polymers, and their chemical interactions in the mucoadhesive suspensions, FTIR analysis was conducted in the present study. This knowledge is essential to produce stable mucoadhesive suspensions without hampering the C-F bond of the quinolone nucleus, which is responsible for the antibacterial activity of the drug..

## MATERIALS AND METHODS

### Materials

The following materials were used for the study: Ofloxacin was obtained from Dr. Reddy's Lab, Hyderabad, India, as a gift sample. Hydroxypropyl methylcellulose (HPMC E15 LV Premium) was supplied by Loba Chemie Pvt. Ltd., India. It was having methoxy group (23.8%) and hydroxypropoxy group (8.3%). Pluronic F 68 and Soya lecithin were purchased from Himedia Laboratories Pvt. Ltd., India. C934, C940, Glycerol, Methyl paraben sodium, Propyl paraben sodium, Sorbitol solution I.P. and Sucrose were supplied by Cosmo Chem. Laboratory, Pune, India. Tri-sodium citrate dehydrate purified was obtained from Merck Specialities Private Limited, Mumbai, India. Ultra pure water was obtained from a Millipore Milli-Q UV water filtration system.

### Methods

#### Preparation of Formulation

##### Preparation of Bulk A

In a beaker, 6 ml water was heated up to 80° C. Then sucrose (10 gm) was added to it under continuous stirring. The temperature was monitored in such a way so that it should not fall below 70° C, till the sucrose was completely dissolved. The prepared syrup was cooled

properly at room temperature and kept overnight. Syrup was filtered using 120 mesh nylon cloth.

#### Preparation of Bulk B

Five millilitre of Ultra pure water was taken in a beaker to which 1.8 ml of sorbitol solution and 0.2 ml glycerin were added. The mixture was stirred properly. To this solution, pluronic F 68 (5%), soya lecithin (1%) and C934/C940/HPMC (5%) in w/w of drug were added with continuous stirring.

#### Preparation of Mucoadhesive Suspension and Ultrasonication

Five millilitre of water was taken in another beaker to which 250 mg of Ofloxacin was added. To the drug suspension, the bulk B and bulk A were added with continuous stirring. Methyl paraben sodium (0.015%w/v) and Propyl paraben sodium (0.08%w/v) were added as preservatives. The volume was made up to 25 ml by Ultra pure water. The  $p^H$  was adjusted by adding citrate buffer (0.75M) to  $p^H$  5.5. Homogenization was carried out for at least 20 min by ULTRASONIC HOMOZENIZER LABSONIC<sup>R</sup> M (SARTORIUS), having operating frequency 30 KHZ and line voltage 230 V/50 HZ, using the probe made up of Titanium of diameter 7 mm and length 80 mm. The setting knob "cycle" was adjusted to 0.8, indicating sound was emitted for 0.8 s and paused for 0.2 s. In this manner, we could expose our sample with 100% amplitude, while reducing the heating effect to 80%. This LABSONIC<sup>R</sup> M generates longitudinal mechanical vibrations with a frequency of 30,000 oscillations / s (30 KHZ). The probes bolted to the sound transducer were made of high-strength Titanium alloys, built as  $\lambda/2$  oscillators. It amplified the vertical oscillation, and transferred the ultrasonic energy via its front surface with extremely high power density into the sample that was to be subjected to ultrasonic waves. In our study, stress applied was sound wave and in addition, mild rise in temperature of the sample occurred during ultrasonication which helped in the homogenization of the suspension.

#### Fourier Transform Infrared Spectroscopy-

After ultrasonication, the polymeric suspension was sprayed on to an aluminum slip with the aid of an atomizer. The fine droplets were dried overnight at room temperature and the solid samples were then collected and powdered. This powder sample was used for FTIR analysis. The Fourier transform infrared analysis was conducted to verify the possibility of interaction of chemical bonds between drug and polymer. FTIR analysis was performed by FTIR Spectrophotometer interfaced with infrared (IR) microscope operated in reflectance mode. The microscope was equipped with a video camera, a liquid Nitrogen-cooled Mercury Cadmium Telluride (MCT) detector and a computer controlled translation stage, programmable in the x and y directions. Solid powder samples were oven dried at around 30°C, finely crushed, mixed with potassium bromide (1:100 ratio by weight) and pressed at 15000 psig (using a Carver Laboratory Press, Model C, Fred S. carver Inc., WIS 53051) to form disc. The detector was purged carefully using clean dry nitrogen gas to increase the signal level and reduce moisture. The spectra were collected in the 400  $cm^{-1}$  to 4000  $cm^{-1}$  region with 8  $cm^{-1}$  resolution, 60 scans

and beam spot size of 10  $\mu\text{m}$ -100  $\mu\text{m}$  [17-19]. The FTIR imaging in the present investigation was carried out using a Perkin Elmer Spectrum RX.

## RESULTS

In FTIR spectra of Ofloxacin, one prominent characteristic peak was found between 3050 and 3000  $\text{cm}^{-1}$ , which was assigned to stretching vibration of OH group and intramolecular hydrogen bonding (Figure 4). This band also suggested the NH stretching vibration of the imino-moiety of piperazinyl groups which was less prominent due to intense OH stretching vibration. The peak at 2700  $\text{cm}^{-1}$  was assigned to  $\nu\text{CH}_3$  of methyl group. The band at 1750-1700  $\text{cm}^{-1}$  represented the acidic carbonyl C=O stretching i.e.,  $\nu_{\text{C=O}}$  [20]. The peak at 1650 to 1600  $\text{cm}^{-1}$  was assigned to  $\nu\text{N-H}$  bending vibration of quinolones. The 1550 to 1500  $\text{cm}^{-1}$  represented the  $\nu\text{CH}_2$  of the aromatic ring. The band at 1450-1400  $\text{cm}^{-1}$  was assigned to the stretching vibration of  $\text{CH}_2$ , confirming the presence of methylene group in benzoxazine ring. The peak at 1400-1350  $\text{cm}^{-1}$  represented the bending vibration of hydroxyl group. The band at 1250 to 1200  $\text{cm}^{-1}$  suggested the stretching vibration of oxo group. In addition, a strong absorption peak between 1050 and 1000  $\text{cm}^{-1}$  was assigned to C-F group. The band at 900-800  $\text{cm}^{-1}$  represented the out of plane bending vibration of double bonded 'enes' or =CH groups (Table 1) [17, 18, 21, 22].

**Table 1: Prominent FTIR Peaks of Ofloxacin**

PEAK ( $\text{cm}^{-1}$ )	GROUP	PEAK ASSIGNMENT
3050-3000	Hydroxyl group	O-H stretching vibration, intramolecular H-bonded
3000-2950	Aromatic, cyclic enes	$\nu=\text{CH}$ & Ar-H
2750	Alkyl groups	$\nu\text{CH}_3$
1750-1700	C=O group of acids	$\nu\text{C=O}$ stretching vibration
1650-1600	Quinolines	$\delta\text{N-H}$ bending vibration
1550-1500	Alkyl groups	$\nu\text{CH}_3$ and $\nu\text{CH}_2$
1450-1400	Methylene group in Benzoxazine	stretching vibration of $\text{CH}_2$
1400-1350	Hydroxyl group	$\delta\text{O-H}$ bending vibration
1250-1200	Oxo group	C-O-C stretching vibration
1050-1000	C-F group	C-F stretching
950-800	Aromatics & enes	=C-H out of plane bending vibration

In case of C934, the FTIR spectra having peak between 3000-2950  $\text{cm}^{-1}$  represented OH stretching vibration, i.e.,  $\nu_{\text{O-H}}$  and intramolecular hydrogen bonds (Figure 5). The prominent peak between 1750 to 1700  $\text{cm}^{-1}$  was assigned to carbonyl C=O stretching band i.e.,  $\nu_{\text{C=O}}$ . The peak at 1250 to 1200  $\text{cm}^{-1}$  represented  $\nu_{\text{C-O-C}}$  for acrylates [17,18]. The ethereal cross linking, was proved by prominent peak at 1160  $\text{cm}^{-1}$ , indicated stretching vibration of  $\nu_{\text{C-O-C}}$  group. The peak at 1450 to 1400  $\text{cm}^{-1}$  was assigned to  $\nu_{\text{C=O}}$  /  $\delta_{\text{O-H}}$  and between 850 and 800  $\text{cm}^{-1}$  was for out of plane bending of C=CH i.e.,  $\delta_{\text{C-H}}$  (Table 2a) [17,18].

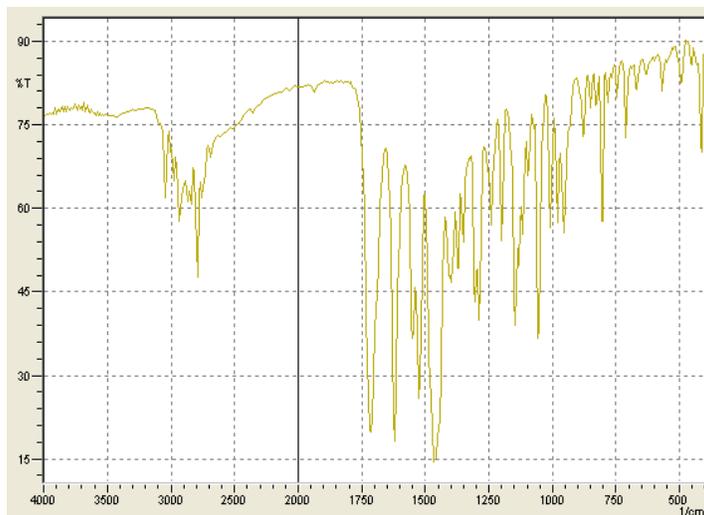


Figure 4: FTIR Spectra of Ofloxacin

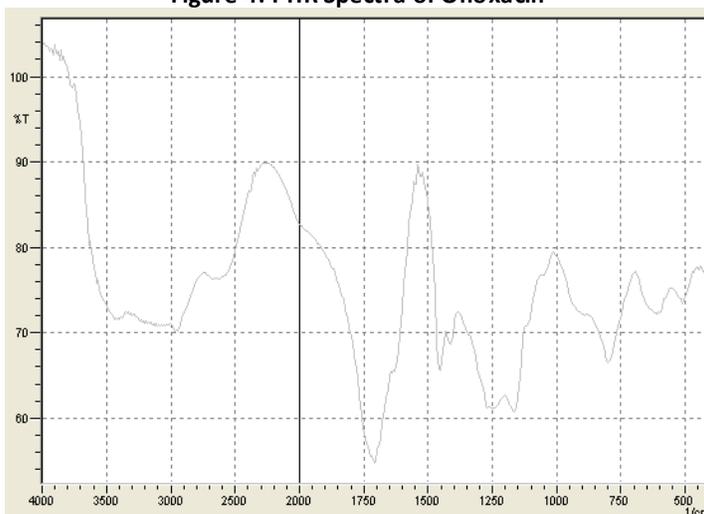


Figure 5: FTIR Spectra of C934

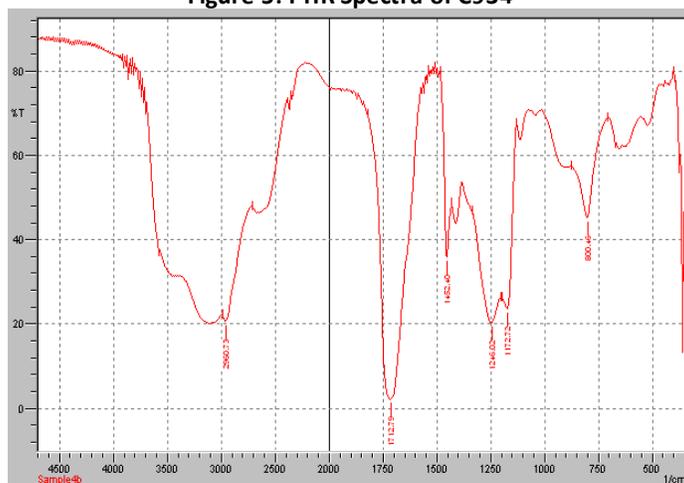


Figure 6: FTIR Spectra of C940

In case of FTIR spectra of C940, similar peaks were found (Figure 6). The FTIR band at  $2960.73\text{ cm}^{-1}$  was assigned to  $\nu_{\text{O-H}}$  i.e., intermolecular hydrogen bonding. While the peak at  $1712.79\text{ cm}^{-1}$  represented  $\nu_{\text{C=O}}$ , the bands at  $1452.40\text{ cm}^{-1}$  and  $1246.02\text{ cm}^{-1}$  were assigned to  $\nu_{\text{C-O}} / \delta_{\text{O-H}}$  and  $\nu_{\text{C-O-C}}$  (for acrylates), respectively. The ethereal cross linking, proved by prominent peak at  $1172.72\text{ cm}^{-1}$ , indicated stretching vibration of  $\nu_{\text{C-O-C}}$  group and finally the band at  $800.46\text{ cm}^{-1}$  was assigned to  $\delta_{\text{C-H}}$  i.e., out of plane bending of C=CH group (Table 2b) [17,18].



Figure 7: FTIR Spectra of HPMC

From FTIR spectra of HPMC, it was found that the peak at  $3500\text{ to }3400\text{ cm}^{-1}$  which indicated OH vibrational stretching (Figure 7) [17,18]. The symmetric stretching mode of  $\nu_{\text{s}}\text{Me}$  and  $\nu_{\text{s}}\text{hydroxypropyl}$  groups was found at  $2900\text{ cm}^{-1}$  in which all the C-H bonds extend and contract in phase [18]. The peak at  $2550\text{--}2500\text{ cm}^{-1}$  was assigned to OH stretching vibration, i.e.,  $\nu_{\text{O-H}}$  and intramolecular hydrogen bonding [17,18]. The band between  $1650\text{ and }1600\text{ cm}^{-1}$  indicated the presence of stretching vibration of  $\nu_{\text{C-O}}$  for six membered cyclic rings. Two bending vibrations might occur within a methyl group. Firstly, the symmetric bending vibration of  $\delta_{\text{s}}\text{Me}$  was involved the in-phase bending of the C-H bonds. Secondly, the asymmetric bending mode of  $\delta_{\text{as}}\text{Me}$  was due to out-of-phase bending of the C-H bonds. While the asymmetric bending vibrations of the methoxy group appeared in the region of  $1500\text{--}1450\text{ cm}^{-1}$ , the symmetric vibrations were mostly displayed in the range of  $1400\text{--}1350\text{ cm}^{-1}$  [23,24]. The band between  $1400\text{ and }1350\text{ cm}^{-1}$  suggested  $\nu_{\text{C-O-C}}$  of cyclic anhydrides. The peak at  $1300\text{--}1250\text{ cm}^{-1}$  was due to  $\nu_{\text{C-O-C}}$  cyclic epoxide. The band at  $1100\text{--}1000\text{ cm}^{-1}$  was for stretching vibration of ethereal C-O-C groups. The peak at  $1000\text{--}950\text{ cm}^{-1}$  was due to  $\nu_{\text{as}}$  of pyranose [25]. The rocking mode of  $\text{CH}_2$  was found in the range of  $850\text{--}800\text{ cm}^{-1}$  (Table 2c) [23]. The computed frequencies of HPMC were in a good agreement with experimental frequencies for carbohydrate region as well as OH and CH regions.

Table 2: Prominent FTIR Peaks of Polymers

a) Prominent FTIR Peaks of C934		
PEAK (cm <sup>-1</sup> )	GROUP	PEAK ASSIGNMENT
3000-2950	Hydroxyl group	O-H stretching vibration, intramolecular H-bonded
1750-1700	C=O group of acids	$\nu_{C=O}$ stretching vibration
1450-1400	Carbonyl group of acids	$\nu_{C=O}$
1250-1200	Acrylates	C-O-C stretching vibration
1160	Ethereal C-O-C group	Stretching vibration of C-O-C group
850-800	Aromatics & enes	=C-H out of plane bending vibration
b) Prominent FTIR Peaks of C940		
2960.73	Hydroxyl group	O-H stretching vibration, intramolecular H-bonded
1712.79	C=O group of acids	$\nu_{C=O}$ stretching vibration
1452.40	Carbonyl group of acids	$\nu_{C=O}$
1246.02	Acrylates	C-O-C stretching vibration
1172.72	Ethereal C-O-C group	Stretching vibration of C-O-C group
800.46	Aromatics & enes	=C-H out of plane bending vibration
c) Prominent FTIR Peaks of HPMC		
3500-3400	Hydroxyl group	O-H stretching vibration, intermolecular H-bonding
2900	Methyl and hydroxypropyl group	$\nu_{C-H}$ stretching of methyl and propyl group
2550-2500	Hydroxyl group	O-H stretching vibration, intramolecular H-bonding
1650-1600	Six membered cyclic	$\nu_{C=O}$
1500-1450	$\delta_{CH}$ , $\delta_{OCH}$ , $\delta_{CCH}$	Assymmetric bending vibration of methyl group in CH <sub>3</sub> O
1400-1350	Cyclic anhydrides	$\nu_{C-O-C}$ and symmetric bending of methoxy group
1300-1250	epoxides	cyclic $\nu_{C-O-C}$
1100-1000	Ethereal C-O-C group	Stretching vibration of C-O-C group
1000-950	Pyranose ring	$\nu_{as}$ of pyranose ring
850-800	CH <sub>2</sub> group	rocking mode of CH <sub>2</sub> group

In the FTIR spectra of the mucoadhesive suspension containing Oflox and C934, the prominent band found between 3550 and 3500 cm<sup>-1</sup> was assigned to  $\nu_{O-H}$ , which was due to single bridge hydrogen bonding (Figure 8). While the peak from 3450 to 3400 cm<sup>-1</sup> was assigned to polymeric  $\nu_{O-H}$  and hydrogen bonding, the band between 2650 and 2600 cm<sup>-1</sup> represented the  $\nu_{O-H}$  i.e., strong hydrogen bonding. The band from 1650 to 1600 cm<sup>-1</sup> was assigned to  $\nu_{C=O}$  i.e., carbonyl stretching vibration. A prominent peak at 1450 cm<sup>-1</sup>(w) was for  $\nu_{C=O}$  /  $\delta_{O-H}$ . The band from 1300 to 1250 cm<sup>-1</sup> was assigned to  $\nu_{C-O-C}$  of acrylates. The peak between 1100 and 1000 cm<sup>-1</sup> represented  $\nu_{C-F}$  groups (Table 3a) [17,18].

In case of FTIR spectra of Oflox with C940, the prominent peak found at 3500-3400  $\text{cm}^{-1}$  was assigned to polymeric  $\nu_{\text{O-H}}$  group (Figure 9). The band between 3100 to 3000  $\text{cm}^{-1}$  represented  $\nu_{\text{C-H}}$  (m). While the peak at 2800-2700  $\text{cm}^{-1}$  suggested intermolecular hydrogen bonding, the band at 1750-1700  $\text{cm}^{-1}$  was assigned to  $\nu_{\text{C=O}}$ . Moreover, the bands at 1650-1600  $\text{cm}^{-1}$  and 1500-1400  $\text{cm}^{-1}$  represented both asymmetric and symmetric stretching vibration of O-C-O group of carboxylic acids, respectively. The peak at 1250-1200  $\text{cm}^{-1}$  indicated  $\nu_{\text{C-O-C}}$  of acrylates and ethers. In addition, the band at 1050-1000  $\text{cm}^{-1}$  was assigned to  $\nu_{\text{C-F}}$  and at 800  $\text{cm}^{-1}$  was for bending vibration of Ar-H groups (Table 3b) [17,18].

In case of FTIR spectra of Oflox with HPMC, the peak from 3100 to 3000  $\text{cm}^{-1}$  was assigned to polymeric  $\nu_{\text{O-H}}$  and hydrogen bonding, the band between 3000 and 2600  $\text{cm}^{-1}$  represented the stretching vibration of  $\nu_{\text{O-H}}$  i.e., strong intermolecular hydrogen bonding (Figure 10). The band from 1650 to 1600  $\text{cm}^{-1}$  was assigned to  $\nu_{\text{C=O}}$  i.e., carbonyl stretching vibration. A prominent peak at 1500-1450  $\text{cm}^{-1}$  (w) was for  $\nu_{\text{C-O}} / \delta_{\text{O-H}}$ . The band from 1400-1350  $\text{cm}^{-1}$  was assigned to  $\delta_{\text{C-O-C}}$ , representing esters and symmetric bending of methoxy groups. The peak between 1100 and 1000  $\text{cm}^{-1}$  indicated  $\nu_{\text{C-F}}$  groups [17, 18]. The band at 1000-950  $\text{cm}^{-1}$  was assigned to  $\nu_{\text{as}}$  of pyranose ring of HPMC [25] (Table 3c).

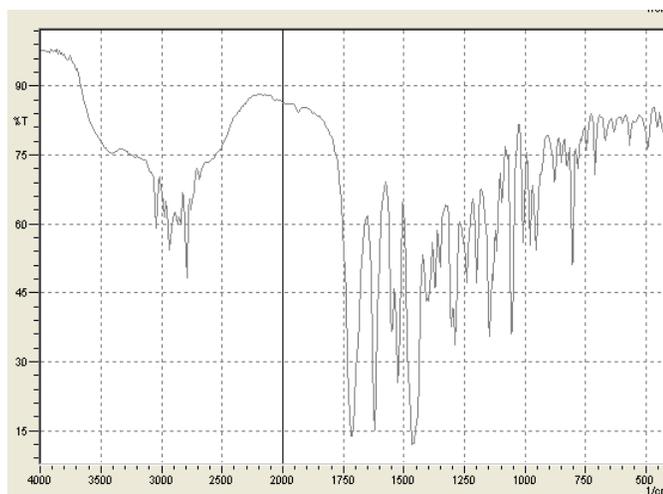


Figure 8: FTIR Spectra of Polymeric Suspension containing Oflox and C934

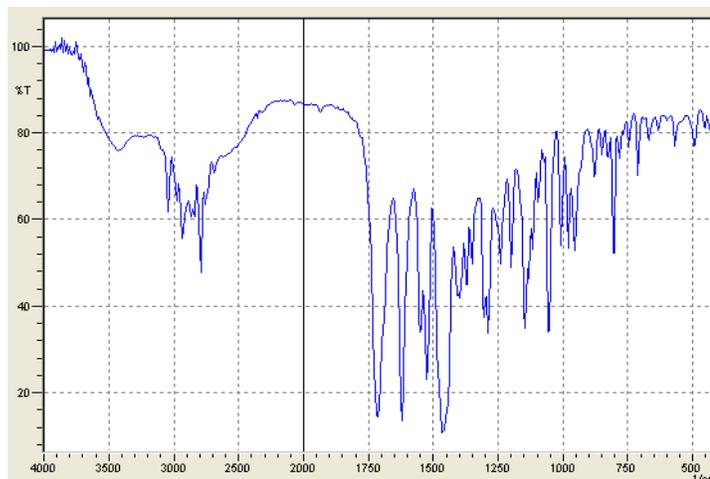


Figure 9: FTIR Spectra of Polymeric Suspension containing Oflox and C940

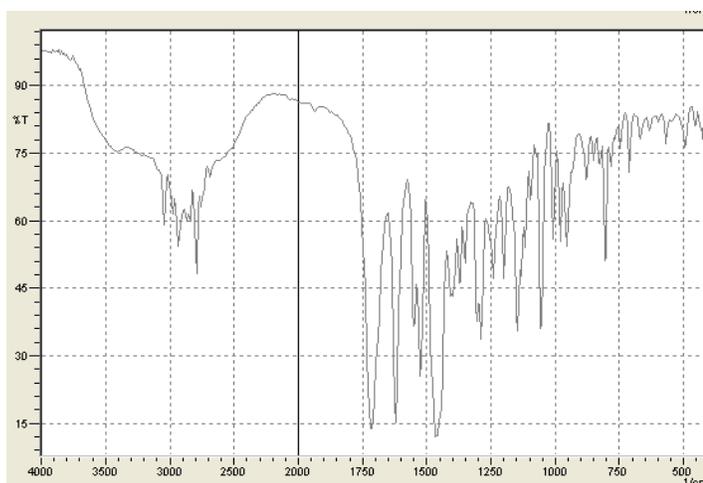


Figure 10: FTIR Spectra of Polymeric Suspension containing Oflox and HPMC

**Table 3: Prominent FTIR Peaks of Ofloxacin Polymeric Suspensions**

<b>a) Polymeric Suspension containing Oflox and C934</b>		
<b>PEAK (cm<sup>-1</sup>)</b>	<b>GROU P</b>	<b>PEAK ASSIGNMENT</b>
3550-3500	Hydroxyl group	H –bonding by single bridge
3450-3400	Polymeric OH groups	$\nu_{O-H}$ , H-bonding
2650-2600	Strong H- bonding	O-H stretching vibration
1650-1600	O-C-O group of acid	$\nu_{as}$ stretching vibration of carbonyl group
1450	O-C-O group of acid	$\nu_s$ stretching vibration of carbonyl group, $\nu_{C-O} / \delta_{O-H}$
1300-1250	Acrylates & esters	C-O-C stretching vibration
1100-1000	C-F groups	$\nu_{C-F}$
800	Aromatic m - distribution	$\delta_{Ar-H}$
<b>b) Polymeric Suspension containing Oflox and C940</b>		
3500-3400	Hydroxyl group	$\nu_{O-H}$
3100-3000	enes	$\nu_{=C-H(m)}$
2800-2700	O-H groups	Intermolecular H-bonded
1750-1700	C=O groups	$\nu_{C=O}$
1650-1600	O-C-O group of acid	$\nu_{as}$ stretching vibration
1500-1450	O-C-O group of acid	$\nu_s$ stretching vibration
1250-1200	Acrylates & esters	C-O-C stretching vibration
1050-1000	C-F groups	$\nu_{C-F}$
800	Aromatic & enes	$\delta_{Ar-H}$ & $\delta_{=C-H}$
<b>c) Polymeric Suspension containing Oflox and HPMC</b>		
3100-3000	Hydroxyl group	O-H stretching vibration, polymeric H-bonded
3000- 2600	Hydroxyl group	O-H stretching vibration, intramolecular H-bonded
1650-1600	O-C-O group of acids	$\nu_{as}$ stretching vibration of acids
1500-1450	O-C-O group of acids	$\nu_s$ stretching vibration of acids, $\nu_{C-O} / \delta_{O-H}$
1400-1350	Esters and Methoxy groups	$\delta_{C-O-C}$ symmetric bending of esters and methoxy groups
1100-1000	C-F group	C-F stretching of Ofloxacin
1000-950	Pyranose ring	$\nu_{as}$ of pyranose ring of HPMC

## DISCUSSION

Infrared (IR) absorption of the functional groups may vary over a wide range. However, it has been found that many functional groups give characteristic IR absorptions at specific narrow frequency ranges [17, 18].

In case of FTIR spectra of Oflox, prominent peaks for  $\nu_{C-O} / \delta_{O-H}$  and  $\nu_{C=O}$  indicated the presence of –CO-, -CHO and -COOH groups (Fig. 4). The presence of above groups can be confirmed by fermi resonance bands for –CHO;  $\nu_{C-O-C}$  bands for esters; and absence of these two for ketones. This suggested the existence of –COOH group in Oflox (Table 1).

In case of FTIR spectra of Carbopol polymers, there were prominent peaks for intramolecular hydrogen bonding,  $\nu_{\text{OH}}$  stretching vibration, carbonylic C=O and C-O stretching vibration, and stretching vibration for the C-O-C, which confirmed the presence of acrylates (Figures 5 and 6). The peak for out of plane bending vibration of =C-H was found between 850 and 800  $\text{cm}^{-1}$  (Tables 2a and 2b). On the other hand, from FTIR spectral analysis of HPMC, it was found that there were both intramolecular and intermolecular hydrogen bondings. In addition, the presence of pyranose ring of  $\beta$  D-glucose monomers was confirmed. The stretching vibration of the cyclic anhydride, methoxy and hydroxypropoxy groups along with epoxide helped in the identification of HPMC (Table 2c) [17, 18, 23, 24, 25].

While comparing the FTIR spectra among the pure Oflox and polymers like C934, C940, HPMC, and the suspensions containing both Oflox and polymers, it was clear that the band position of C=O group was affected by esterification and conjugation involving C=O group. Here, the stretching vibration of C=O in pure Oflox was found from 1750 to 1700  $\text{cm}^{-1}$  which was lowered to 1650-1600  $\text{cm}^{-1}$  in the formulations might be due to formation of  $\beta$ -ketoesters (Figures 4 and 8-10). The FTIR peaks assigned to  $\nu_{\text{C-O}}$  and  $\nu_{\text{C-O-C}}$  representing acrylates and esters confirmed the esterification between polymeric -OH and -COOH groups of drug (Oflox). The stretching vibration of C-F group of the drug remained nearly unaltered which indicated that the antibacterial activity of the drug was not affected appreciably in different suspensions. Another probability of interaction was hydrogen bonding i.e., intermolecular hydrogen bonding due to prominent FTIR peaks between 3550 and 3500  $\text{cm}^{-1}$ , 3450 and 3400  $\text{cm}^{-1}$ , and 2650 and 2600  $\text{cm}^{-1}$  represented single bridge O-H...O, polymeric O-H...O-H...O-H and strong hydrogen bonding, respectively. The hydrogen bonded -OH stretching vibration occurred over a wide range, 3550-2600  $\text{cm}^{-1}$ . In case of intramolecular hydrogen bonding, FTIR bands were sharp while in intermolecular hydrogen bonding they were broad. However, it was less broad than which was required for chelation. The bending vibration of O-H group gave medium to strong bands in the region around 1450  $\text{cm}^{-1}$ . The FTIR peak at 800  $\text{cm}^{-1}$  suggested the probability of out of plane bending of =ene bond and m-substitution of  $\delta_{\text{Ar-H}}$  hydrogen atom (Tables 1 and 3) [17,18]. The C=O group of drug lowered the stretching vibration of C=O frequency indicating deprotonation and probably interaction of the said carboxylic C=O moiety with the polymers. However, a definitive conclusion about the keto group in the bonding to the polymer could be deduced because the corresponding band found from 1650 to 1600  $\text{cm}^{-1}$  was due to probability of the formation of  $\beta$ -ketoesters [26]. From the above data, it can be inferred that the carboxylic group of Oflox undergoes the interaction with the polymer, as would be expected chemically. Thus the nitrogen atoms aren't likely to be involved in binding or the interaction. The nitrogen atom of the quinolone ring, 1-ortho to fluorine, is less electron rich due to electron deficient fluoroquinolone ring. In addition, cyclopropyl and piperazinyl groups sterically hinder the reaction. The possibility of involvement of imino moiety of the piperazinyl group is also less prominent due to intense OH stretching vibration. The bands in the region of 3500-2700  $\text{cm}^{-1}$  could be assigned to the asymmetric and symmetric stretching vibrations of the OH groups of the inner and outer sphere of polymers. The shift in the characteristic bands of the FTIR spectra suggests change in their intensity leading to the appearance of several absorbance bands of the asymmetric and symmetric stretching vibrations and overtone of the deformation vibrations. This confirms the presence of the hydrogen bonds [17,18,26]. By

comparing the FTIR spectra among the pure drug, polymers and the suspensions containing drug and polymer, the FTIR peak of Oflox from 1750 to 1700  $\text{cm}^{-1}$  was not detected in the formulations probably due to interaction with polymers. The missing peak was replaced with two very strong characteristic bands, in the range of 1650-1600  $\text{cm}^{-1}$  and at 1450  $\text{cm}^{-1}$ , were assigned to  $\nu_{(\text{O}-\text{C}-\text{O})}$  asymmetric and symmetric stretching vibrations, respectively [25,27]. The difference  $\Delta [\nu_{(\text{CO}_2)_{\text{asym}}}-\nu_{(\text{CO}_2)_{\text{sym}}}]$  is a useful characteristic for determining the involvement of the carboxylic group of Oflox. The  $\Delta$  value for the interaction falls in the range of 183 - 250  $\text{cm}^{-1}$ , indicating the deprotonation of the carboxylic acid group and interaction between drug and polymers (Table 1 and 3) [27].

## CONCLUSIONS

On the basis of our interpretation, it can be concluded that by preparing mucoadhesive suspensions of Ofloxacin with these three polymers following a novel method of ultrasonication, there is a very good interaction between the carboxylic group of drug and hydroxyl group of polymers. This leads to esterification and intermolecular hydrogen bonding, by virtue of which stable mucoadhesive suspensions could be produced without hampering the C-F bond of the quinolone nucleus, which is responsible for the antibacterial activity of the drug. As a result, these polymers may be considered as effective carriers for Ofloxacin.

## REFERENCES

- [1] Garg R, Gupta GD. Trop J Pharm Res 2008; 7(3): 1055-66.
- [2] Hosmani AH. Carbopol and its Pharmaceutical Significance: A Review;
- [3] [cited 2010 Jan 20]. Available from: <http://www.pharmainfo.net/reviews/carbopol-and-its-pharmaceutical-significance-review>.
- [4] Cruz AP, Rodrigues PO, Cardoso TM, Silva MAS. Am J Pharm 2007; 26 (2): 171-78.
- [6] Bettini R, Colombo P, Peppas NA. J Control Release 1995; 37(1-2): 105-11.
- [7] Qiu Y, Park K. Adv Drug Delivery Rev 2001; 53(3): 321-339.
- [8] Bromberg LE, Ron ES. Adv Drug Delivery Rev 1998; 31: 197-21.
- [9] Galaev IY, Mattiasso B. Trends Biotechnol 1999; 17(8): 335-40.
- [10] Jeong B, Gutowska A. Trends Biotechnol 2001; 20: 305-11.
- [11] Gupta P, Vermani K, Garg S. Drug Discovery Today 2002; 7: 569-79.
- [12] Yoshida R, Sakai K, Okana T, Sakurai Y. Adv Drug Delivery Rev 1993; 11: 85-108.
- [13] Fatimi A, Tassin JF, Quillard S, Axelos MAV, Weiss P. Biomater 2008; 29(5): 533-543.
- [14] Siepmann J, Peppas NA. Adv Drug Deliv Rev 2001; 48: 139-57.
- [15] Phaechamud T. AAPS Pharm Sci Tech. 2008; 9(2): 668- 674.
- [16] Talukdar MM, Michoel A, Rombout P, Kinget R. Int J Pharm 1996; 129: 233-41.
- [17] Katiknani PR, Upadrashta SM, Neau SH, Mitra AK. Int J Pharm 1995; 123: 119-25.
- [18] Gao P, Skoug JW, Nixon PR, Ju RT, Stemm NL, Sung K. J Pharm Sci 1996; 85(7): 732-40.
- [19] Silverstein RM, Webster FX. Spectrometric Identification of Organic Compounds. 6<sup>th</sup> Ed. New York, Jhon Wiley and Sons. 2002, pp.71-109.

- [20] Dani VR. Organic Spectroscopy. 1<sup>st</sup> Ed. New Delhi, Tata McGraw-Hill Publishing Company Limited. 1995, pp.86-168.
- [21] Precautions for Making KBr Pellets; Available from [http://www.chemistry.nmsu.edu/Instrumentation/KBr\\_New.html](http://www.chemistry.nmsu.edu/Instrumentation/KBr_New.html), accessed on 20.01.2010.
- [22] Kulkarni PV, Keshavayya J. Int J Pharm Sci 2010; 2(2): 77-82.
- [23] Anam AA, Fandi Z, Gryta M, Balcerowiak W. Pak J Appl Sci 2002; 2(10): 940-44.
- [24] Pandya SJ, Bhalekar MR, Harinarayana D, Shah SS, Darji D. Int J Pharma Res 2010; 2(3): 28-32.
- [25] Raj A, Raju K, Varghese HT, Granadeiro CM, Nogueira HIS, Panicker CY. J Braz Chem Soc 2009; 20(3): 549-559.
- [26] Govindarajan M, Periandy S, Ganesan K. E-Journal Chem 2010; 7(2): 457-64.
- [27] Ibrahim M, Alaam M, El-Haes H, Jalbout AF, de Leon A. Analysis of the structure and vibrational spectra of glucose and fructose. Eclat Quím 2006; 31(3); Available from: [http://www.scielo.br/scielo.php?script=sci\\_arttext&pid=S0100-46702006000300002&lng=en&nrm=iso&tlng=en](http://www.scielo.br/scielo.php?script=sci_arttext&pid=S0100-46702006000300002&lng=en&nrm=iso&tlng=en), accessed on 25.02.2011.
- [28] Garrido NJ, Perello L, Ortiz R, Alzuet G, Alvarez MG, Canton E, Gonzalez ML,
- [29] Granda SG, Priede MP. J Inorg Biochem 2005; 99: 677-89.
- [30] Efthimiadou EK, Psomas G, Sanakis Y, Katsaros N, Karaliota A. J Inorg Biochem 2007; 101: 525-35.